



# APlink AP Broad Detection Kit for Mouse and Rabbit Antibodies

(Alkaline Phosphatase labeled streptavidin-biotin detection system for broad spectrum)

	Catalog No.:	D07-110	110mL 🔀
Storage: 4-8°C		D07-60	60mL
		D07-18F	18mL
		D07-6F	6mL
		D07-18A	18mL
		D07-6A	6mL

#### **Intended Use:**

APlink AP Broad Detection Kit uses biotinylated secondary antibody and Alkaline Phosphatase (AP) labeled-streptavidin to detect mouse and/or rabbit primary antibody (user-supplied) that bind to antigens in human tissue or cell preparations under light microscopy. The most commonly used specimens for this system are: frozen tissue, paraffin-embedded tissue, freshly prepared lymphocytes and fixed culture cells. Alkaline Phosphatase (AP) labeled-streptavidin and biotinylated secondary antibody amplification system has become a standard technique in immunochemical staining<sup>1,2</sup>. APlink AP Broad Detection Kit uses human-absorbed, biotinylated, affinity-purified secondary antibody reacts with the user supplied primary antibody bound to the specific epitope of the antigen in tissue or cell. Alkaline Phosphatase (AP) labeled streptavidin then reacts with biotinylated secondary antibody to form an AP-streptavidin-biotin complex. The AP enzyme of the streptavidin complex catalyzes the substrate/chomogen such as Fast-Red, GBI-Permanent Red, or BCIP/NBT to form a red (Fast-Red or GBI-Permanent Red) or dark blue/purple (BCIP/NBT) color deposit at the antigen site. The antigen then can be visualized under microscope. Compared to traditional ABC methods which uses avidin, APlink AP Broad Detection Kit demonstrates stronger binding strength to bind biotin and less non-specific background staining.

Higher sensitivity and lower background give this kit a higher signal-noise ratio. It also provides users cost effective method for their research. End users may choose Fast-Red, GBI-Permanent Red, or BCIP/NBT chromogen depending on their preferences.

#### **Kit Components:**

### A Kit

Component No.	Content	D07-6A	D07-18A	D07-60	D07-110
Reagent 1	Pre-Block Solution (RTU)	6mL	18mL	60mL	110mL
Reagent 2	Biotinylated anti-Mouse & Rabbit (RTU)	6mL	18mL	60mL	110mL
Reagent 3	Streptavidin-AP (RTU)	6mL	18mL	60mL	110mL
Reagent 4A	GBI-Permanent Red Substrate (RTU)	7mL	18mL	NA	NA
Reagent 4B	GBI-Permanent Red Activator (5x)	1.4mL	3.6mL	NA	NA
Reagent 4C	GBI-Permanent Red Chromogen (100x)	70μL	180μL	NA	NA

## F Kit

Component No.	Content	D07-6F	D07-18F
Reagent 1	Pre-Block Solution (RTU)	6mL	18mL
Reagent 2	Biotinylated anti-Mouse & Rabbit (RTU)	6mL	18mL
Reagent 3	Streptavidin-AP (RTU)	6mL	18mL
Daggard 4A	Fast Red tablets (Tablets)	6 tablets	15
Reagent 4A			Tablets
Reagent 4B	Fast Red Substrate (RTU)	5mLx6	40mlx2

## **Recommended Protocol:**

- 1. Fixation: To ensure the quality of the staining and obtain reproducible performance, the user needs to supply appropriately fixed tissue and well prepared slides.
- 2. Tissue need to be adhered to the slide tightly to avoid tissue falling off.
- 3. Paraffin embedded section must be deparffinized with xylene and rehydrated with a graded series of ethanol before staining.
- Cell smear samples should be made into as thin monolayer as possible to obtain satisfactory results.
- 5. Three control slides will aid the interpretation of the result: positive tissue control, reagent control (slide treated with Isotype control reagent), and negative control.
- 6. Start staining procedures: DO NOT let specimen or tissue dry from this point on.
- 7. We recommend TBS-T to be used as the wash buffer to get the highest sensitivity and clean background. Phosphate in the PBS-T may inhibit the activity of the alkaline phosphatase. Note: 1X TBS-T =50mM Tris HCl, 150mM NaCl, 0.05% Tween-20 pH7.6. GBI sells 10xTBS-T for your convenience (B11xx)

Reagent	Staining Procedures	Incubatio nTime
1. HIER Pretreatment: refer to antibody spec. sheet	<ul> <li>a. Heat Induced Epitope Retrieval (HIER) may be required for primary antibody. Refer to antibody datasheet.</li> <li>b. Wash with PBS-T containing 0.05% Tween-20 or 1X TBS-T(See note 7 above); 3 times for 2</li> </ul>	

	minutes each.	
2. Reagent 1:	a. Add 2 drops or enough volume of <b>Regent 1</b> (Pre-blocking Solution) to completely cover the	
Pre-blocking Solution (RTU)	tissue section and incubate for 10 min. b. Blot off solution. <b>DO NOT RINSE</b> .	1min.
2 Primary antibody:	Note: Investigator needs to optimize dilution and incubation time.	
3. Primary antibody:	a. Apply 2 drops or enough volume of Primary antibody to cover the tissue section completely.	
Supplied by user.	Incubate in moist chamber for 30-60min.	30-60min.
	b. Wash with PBS-T containing 0.05% Tween-20 or <b>1X TBS-T</b> ; 3 times for 2 minutes each.	
4. Reagent 2:	a. Apply 2 drops or enough volume of <b>Reagent 2</b> (Biotinylated anti-Mouse & Rabbit) to cover	
Biotinylated anti-Mouse & Rabbit	the tissue section completely and incubate for 10min.	10min.
	b. Wash with PBS-T containing 0.05% Tween-20 or <b>1X TBS-T</b> ; 3 times for 2 minutes each.	10IIIII.
(RTU)	- Analy 2 days and the laws of <b>D</b> - and 2 (General did AD) to a second did the s	
5. Reagent 3:	a. Apply 2 drops or enough volume of <b>Reagent 3</b> (Streptavidin-AP)to cover the tissue section completely and incubate for 10min.	10 .
Streptavidin-AP (RTU)		10min.
	b. Wash with 1xTBS-T only, 3 times for 2 minutes each.  Refer to manufacture data sheet if chromogen is supplied by user.	
6. <b>Reagent 4:</b> Chromogen:	Recommended protocol for chromogen using our kit:	
Fast-Red, or GBI-Permanent Red (To	1. Fast Red:	
get maximum sensitivity of AP	a. Dissolve one Fast Red tablet into one 5mL substrate buffer. Vortex until tablet is dissolved. It	
polymer, Please repeat chromogen	usually takes 20 minutes to dissolve completely.	
step), or BCIP.NBT	b. Chromogen must be used within 1 hour.	
step), or Bell in Bi	c. Apply 100ul or more Fast-Red solution to completely cover the tissue section and incubate 10	
	minutes at room temperature.	
	d. After proper color development, wash with distill water for 2 minutes, 3 times	
	e. DO NOT Dehydrate tissue after staining. Fast-Red is alcohol soluble.	
	2. GBI-Permanent Red:	
	Note: Shake GBI-Permanent Red Activator before adding into GBI-Permanent Red Substrate.	
	a. Add 200µL of Reagent 7B (Activator) into 1mL of Reagent 7A (Substrate) and mix well. Add	
	10μL of Reagent 7C (Chromogen) into the mixture and mix well.	
	[Note: For fewer slides, Add 100µL of Reagent 7B (Activator) into 500µL of Reagent 7A	
	(Substrate) and mix well. Add 5μL of Reagent 7C (Chromogen) into the mixture and mix	
	well.]  h Apply 2 drops (100uL) or apough volume of CPL Permanent Red working solution to	
	b. Apply 2 drops (100μL) or enough volume of GBI-Permanent Red working solution to completely cover the tissue. Incubate for 10 min, observe appropriate color development. To	
	increase AP signal aspirate or tap off chromogen and apply 2-3 drops (100µL) again of the	
	GBI-Permanent Red working solution to completely cover the tissue for additional 5 to	
	10min.	
	c. Rinse well with distilled water.	
	3. BCIP/NBT: order separately, Cat. No. C05-100 or C05-18	
	a. Add two drops (about 100ul) of Ready-to-use BCIP/NBT to cover the tissue section for 5-10	
	minutes. Monitor the color development under a microscope.	
	b. Rinse with distill water for 2 minutes, 3 times.	
7. Hematoxylin:	a. Counterstain with 2 drops or enough volume to cover tissue completely and wait about 10-20	
Supplied by user	seconds.	
err	b. Rinse thoroughly under tap water for 1-2 min.	
	c. Put slides in PBS until show blue color (about 30-60 seconds)	
	d. Rinse well in distilled water	
8. Mounting media:	Follow the manufacturer's data sheet procedure for mounting.	
Supplied by user	Recommended product: 1. GB-Mount: Cat. No. E01-18 (18mL) for AEC, Fast-red, GBI-Permanent Red and AP-blue,	
	DAB, BCIP/NBT.	
	2. O-Mount: Cat. No. E02-18 (18mL), for DAB and BCIP/NBT	
	3. Simpo-Mount: Cat. No. E03-18 (18mL), or E03-100 (100mL), universal permanent mounting	
	medium	
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## **Protocol Notes:**

- The fixation, tissue slide thickness, antigen retrieval and primary antibody dilution and incubation time affect results significantly. Investigator needs to consider all factors and determine optimal conditions when interpreting the result.
- 2. Tissue staining is dependent upon the proper handling and processing of tissues prior to staining. Improper tissue preparation may lead to false negative results or inconsistent results.
- 3. Do not mix reagents from different lot.
- 4. Do not allow the slides to dry at any time during staining.

## **Related Products:**

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Product	Catalog No.	Size	Product	Catalog No.	Size
APlink AP Mouse Bulk Kit	D08-110	110mL	Fast Red Kit	C03-60	60mL
APlink AP Mouse Fast Red Kit	D08-18F / D08-6F	18mL / 6mL	AP-Red+ Kit (40x	C04-8	8mL

			concentrate)		
APlink AP Mouse GBI-Permanent Red	D08-18A / D08-	18mL / 6mL	BCIP/NBT Kit	C05-100 / C05-	100mL / 18mL
Kit	6A			18	
APlink AP Rabbit Bulk Kit	D09-110	110mL	GB-Mount (Aqueous)	E01-18	18mL
APlink AP Rabbit Fast Red Kit	D09-18F / D09-6F	18mL / 6mL	O-Mount (Organic)	E02-18	18mL
APlink AP Rabbit GBI-Permanent Red	D09-18A / D09-	18mL / 6mL	Simpo-Mount (Aqueous)	E03-100 / E03-18	100mL / 18mL
Kit	6A				
Streptavidin-AP (RTU)	D29-110 / D29-18	100mL / 18mL	GBI-Permanent Red Kit	C13-18/ C13-120	18mL / 120mL

#### **Precautious:**

Handle all specimens as potential infectious materials, wear gloves and protection cloth.

#### Remarks

For research use only.

#### **References:**

- 1. Elias, J.M. et al. Sensitivity and Detection Efficiency of the Peroxidase antiperoxidase (PAP) Avidin-Biotin Peroxidase Complex (ABC), and Peroxidase-Labeled Avidin-Biotin (LAB Methods. AM J Clin Pathol 92:62-67, 1989.
- 2. Polak, J.M and Van Noorden, S. Introduction to Immunocytochemistry Second Edition. Bios Scientific Publishers. 41-54. 1997.